

The $\alpha 7$ Nicotinic Acetylcholine Receptor in Neuronal Plasticity

Ron S. Broide¹, and Frances M. Leslie^{*,2}

¹ Division of Neuroscience, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030;

² Department of Pharmacology, University of California, Irvine, CA

Abstract

A growing body of evidence indicates that neuronal nicotinic acetylcholine receptors (nAChRs), in addition to promoting fast cholinergic transmission, may modulate other neuronal activities within the central nervous system (CNS). In particular, the $\alpha 7$ nAChR is highly permeable to Ca^{2+} and may serve a distinct role in regulating neuronal plasticity. By elevating intracellular Ca^{2+} levels in discrete neuronal locations, these ligand-gated ion channels may influence numerous physiological processes in developing and adult CNS. In this article, we review evidence that both pre- and postsynaptic $\alpha 7$ nAChRs modulate transmitter release in the brain and periphery through Ca^{2+} -dependent mechanisms. The possible role of $\alpha 7$ nAChRs in regulating neuronal growth and differentiation in developing CNS is also evaluated. We consider an interaction between cholinergic and glutamatergic transmission and propose a hypothesis on the possible coregulation of intracellular Ca^{2+} by N-methyl-D-aspartate (NMDA) receptors and $\alpha 7$ nAChRs. Finally, the clinical significance of alterations in the normal function of $\alpha 7$ nAChRs is discussed as it pertains to prenatal nicotine exposure, schizophrenia, and epilepsy.

Index Entries: α -Bungarotoxin; nicotine; acetylcholine; cholinergic; plasticity; thalamocortical; cortex; trophic factors.

Introduction

Neuronal development and plasticity are influenced greatly by external signals acting as cues for particular cellular responses. In addition to the influence of growth factors in these

processes, there is a growing body of evidence that neurotransmitters may serve as neurotrophic agents in the peripheral (PNS) and central nervous system (CNS; reviewed in 1–3). Studies have shown that neurotransmitters can modulate the levels of a specific

* Author to whom all correspondence and reprint requests should be addressed.

second messenger(s), such as intracellular free Ca^{2+} , and thereby influence various aspects of cellular plasticity (4).

Numerous investigations have indicated a possible role for cholinergic systems in mediating trophic actions in the developing and adult CNS (5–8). Although a great deal of evidence has implicated muscarinic receptors (5,9), there has been relatively little attention paid to the role of nicotinic acetylcholine receptors (nAChRs) in neuronal plasticity. However, an increasing number of studies are demonstrating that nAChRs, in addition to mediating fast cholinergic transmission, may also play a broader role in the development and modulation of neuronal synapses (10). One particular nAChR subtype that has received much attention in recent years is the $\alpha 7$ nAChR. Since its initial cloning (11), followed by an instrumental review (12), which helped propel this receptor into recognition, there has been a plethora of studies attempting to analyze the functional role of $\alpha 7$ nAChRs in the CNS.

This article relates the available evidence on the potential role(s) of $\alpha 7$ nAChRs in neuronal plasticity. It is not intended to be an historical perspective of the $\alpha 7$ nAChR, since such qualified accounts already exist (12–14). We begin by summarizing the molecular, pharmacological, and physiological diversity of neuronal nAChRs, while focusing on the $\alpha 7$ nAChR. Next, we discuss the roles of pre- and postsynaptic $\alpha 7$ nAChRs as they relate to synaptic plasticity. This is followed by an analysis of $\alpha 7$ nAChRs in the developing CNS, along with the cholinergic source(s) for their activation. In addition, we consider the interaction between cholinergic and glutamatergic transmission as it pertains to the $\alpha 7$ nAChR. Finally, we discuss the possible clinical significance of alterations in the normal function of $\alpha 7$ nAChRs. Although there are also numerous studies on the functional roles of other neuronal nAChR subtypes, we focus here specifically on the $\alpha 7$ nAChR. For more comprehensive descriptions of nAChRs, the reader is referred to several excellent reviews (10,14–19).

Background

The $\alpha 7$ nAChR is a member of a large family of neuronal nAChRs found in the vertebrate PNS and CNS. These ligand-gated ion channels are pentamers, generally composed of α and β subunits (2 α and 3 β). To date, 11 nAChR subunits have been identified in vertebrates and designated as either α -type ($\alpha 2$ – $\alpha 9$) or β -type ($\beta 2$ – $\beta 4$) based on their homology to the muscle $\alpha 1$ subunit. The $\alpha 8$ subunit has been found in avians, but not in mammals (20), and the $\alpha 9$ subunit is only expressed in certain endocrine cells and sensory end organs (21). Heterologous expression of nAChR subunits in *Xenopus* oocytes indicates that many (but not all) of the α and β subunits can coassemble to form functional nAChR channels that are distinct in their biophysical and pharmacological properties (14,16). Moreover, expression of $\alpha 7$, $\alpha 8$, or $\alpha 9$ subunits alone gives rise to functional homo-oligomeric, acetylcholine- (ACh) gated channels that are blocked by nanomolar concentrations of the snake toxin, α -bungarotoxin (α -BTX; 20–22).

Whereas data from in vitro expression studies have identified potential nAChR subtype candidates, more recent studies have helped characterize possible subtypes of native neuronal nAChRs in the CNS (17–19). Although physiological and pharmacological studies have confirmed the existence of distinct functional nAChRs in different regions of the brain, they appear to belong to two major classes: α -BTX-insensitive and α -BTX-sensitive. Receptors containing $\alpha 3$, $\alpha 4$, or $\beta 2$ subunits form the bulk of α -BTX-insensitive nAChRs in the vertebrate brain and account for the vast majority of high-affinity nicotinic agonist binding (23–26). On activation by ACh, these ion channels become permeable to Na^+ , K^+ , and Ca^{2+} ions and show a relatively slow decaying inward current (16). The α -BTX-insensitive channels are found in brain regions, such as the nigrostriatal and mesolimbic pathways, where they modulate dopamine release (18,27,28), and mediate the locomotor and reinforcing properties of nicotine (29–31).

Neuronal nAChRs of the α -BTX-sensitive class contain $\alpha 7$ subunits and account for most of the α -BTX binding in the vertebrate brain (32,98). In the chick CNS and PNS, there is strong evidence for heterologous complexes of $\alpha 7$ along with $\alpha 8$ (34,35), and more recently with $\alpha 5$ (36). However, in mammals, $\alpha 7$ nAChRs appear to be predominantly homomeric ion channels (32). These channels have a high relative permeability to calcium and are blocked by nanomolar concentrations of α -BTX (22,37). Furthermore, activation of $\alpha 7$ nAChRs can produce a rapidly decaying inward current that can quickly elevate intracellular levels of free calcium in neurons, either directly through the channel, or indirectly via depolarization and consequent activation of voltage-gated Ca^{2+} channels (39,40,84).

Presynaptic $\alpha 7$ nAChRs

A general problem in addressing nAChR function in the brain has been the inability to demonstrate clearly ACh-evoked synaptic responses from these receptors, and has been aggravated by the lack of specific agonists and antagonists for each of the receptor subtypes (14). However, the functional characterization of nAChRs has improved immensely in recent years because of subtype-selective ligands, as well as advances in physiological recording techniques that have enabled the analysis of nAChR currents expressed in single neurons (16,17,41). Therefore, a growing body of evidence suggests that nAChRs found at presynaptic terminals in the CNS may have an important role in modulating neurotransmitter release (10,18). Because of their high Ca^{2+} permeability and rapid desensitization, $\alpha 7$ nAChRs provide a unique mechanism for cholinergic regulation of transmitter release throughout the brain. Indeed, recent studies have shown that activation of presynaptic $\alpha 7$ -containing nAChRs can enhance and sometimes elicit release of the excitatory amino acid transmitter, glutamate, from synaptic terminals in developing and adult brains through a

calcium-dependent mechanism. These results were obtained in culture and slice preparations from various brain regions expressing high levels of $\alpha 7$ nAChRs, including the chick medial habenula (42), and lateral geniculate nucleus (43), rat hippocampus (41), olfactory bulb (44), and sensory neocortex (45). Thus, $\alpha 7$ nAChRs may be involved in modulating sensory processing and cognitive tasks, such as learning and memory.

By mediating glutamatergic transmission, $\alpha 7$ nAChRs may also indirectly regulate other neurotransmitter systems. Studies in animals have demonstrated that nicotine increases release of dopamine in the nucleus accumbens by activation of nAChRs in the ventral tegmental area (VTA; 29,30,46,47). These nAChRs include presynaptic $\alpha 7$ nAChRs located on glutamatergic afferents in the VTA, which increase glutamate release and, in turn, stimulate mesolimbic dopamine neurons (48). These results indicate that $\alpha 7$ nAChRs may also have an important role in modulating the reinforcing effect of natural rewards as well as that of various drugs of abuse, such as nicotine.

In addition to facilitating glutamate release, activation of $\alpha 7$ -containing nAChRs on presynaptic sites can potentiate the release of several other neurotransmitters. A recent study in rat brainstem slices has demonstrated nicotine-stimulated release of norepinephrine from dorsal raphe neurons (49). The effect on norepinephrine release was calcium-dependent and inhibited by methyllycaconitine. Although this suggests the presence of presynaptic $\alpha 7$ -containing nAChRs, the high concentration of methyllycaconitine used in this study may also indicate the involvement of other nAChR subtypes. Activation of presynaptic $\alpha 7$ nAChRs in PNS ganglion neurons has been shown to stimulate ACh release (42,50), presumably acting as a positive feedback mechanism for cholinergic transmission. The potentiation of γ -aminobutyric acid (GABA) release from hippocampal interneurons by presynaptic $\alpha 7$ nAChRs has also been proposed (51). However, since most of the $\alpha 7$ nAChRs in interneurons are believed to play a postsynaptic role (*see* Postsynaptic $\alpha 7$

nAChRs; refs. 52–55), such findings are still preliminary. Nevertheless, these data indicate that a major role for presynaptic $\alpha 7$ nAChRs in the vertebrate CNS is to modulate neurotransmitter release and thereby contribute to cholinergic modification of information processing throughout the brain. Further studies will most likely focus on the secondary mechanism(s) involved in this process.

Postsynaptic $\alpha 7$ nAChRs

Although cholinergic transmission mediated by postsynaptic nAChRs is well established in the periphery, it has been considerably more difficult to demonstrate these nicotinic responses in the CNS (10). The $\alpha 7$ neuronal nAChR has been especially elusive in this regard, presumably because of its fast-desensitizing currents and low agonist affinity. However, with recent advances in recording techniques that make it possible to detect synaptic events on a faster time scale, an increasing number of laboratories have now demonstrated evoked synaptic currents in both PNS and CNS neurons, mediated by post- and perisynaptic $\alpha 7$ nAChRs. By general definition, these postsynaptic responses were evoked under pharmacologically isolated conditions by application of specific receptor agonists and blocked by selective receptor antagonists.

Initial studies of chick ciliary ganglion neurons have shown that these cells express a large number of $\alpha 7$ -containing nAChRs (56). Despite their perisynaptic localization (57,58), $\alpha 7$ -containing nAChRs have been shown to produce a large fraction of the synaptic currents within these ganglion neurons (59–61). In the rat brain, fast cholinergic transmission mediated by postsynaptic $\alpha 7$ nAChRs has recently been observed within the hippocampus (52–55), and the olfactory bulb (44), and associated with modulation of GABA release. These results are the first to indicate that $\alpha 7$ nAChRs may also participate in mediating cholinergic neurotransmission in the brain. A

more recent study, however, suggests that the GABAergic circuitry in the hippocampus may be slightly more complicated, consisting of inhibitory and disinhibitory responses that are modulated by nicotinic receptors at both pre- and postterminal regions (62). Thus, neuroanatomical investigations may be required to help determine the precise cellular localization of $\alpha 7$ nAChRs in this area of the brain.

All together, these studies suggest that pre- and postsynaptic $\alpha 7$ nAChRs can play significant roles in cholinergic modulation of transmitter release in the CNS by mediating the release of both excitatory and inhibitory signals through Ca^{2+} -dependent mechanisms. This would have important implications for neuronal plasticity. For example, because of their capacity to regulate neurotransmission in the hippocampus, $\alpha 7$ nAChRs may play a role in modulating the induction of long-term potentiation. Such a role has been suggested by several studies (63,64) and is consistent with the proposed functions of $\alpha 7$ nAChRs in the hippocampus (17). Furthermore, $\alpha 7$ nAChRs may be involved in cholinergic facilitated reorganization of cortical maps in the adult brain (65–68).

$\alpha 7$ nAChRs in the Developing CNS

In addition to their role in modulating synaptic neurotransmission, $\alpha 7$ nAChRs may play an important role in regulating neuronal growth and differentiation. This hypothesis is supported by three separate, but related lines of evidence. First, several studies have demonstrated that $\alpha 7$ nAChR expression is highly regulated in the developing brain during a critical period of synaptic plasticity. In both the chick and rodent, there is strong expression of $\alpha 7$ mRNA and protein in embryonic brain (11,69,70), with a subsequent reduction in many brain regions during postnatal development (70–73). Within the rodent neocortex, for example, transient $\alpha 7$ mRNA and protein expressions delineate sensory regions, with a spatiotemporal correspondence to ingrowing thalamic afferents.

Moreover, this increased $\alpha 7$ nAChR expression is tightly regulated by thalamic afferent activity (74). Additionally, other factors associated with neuronal activity, such as neurotrophic factors (75), protein kinases (76,77), arachidonic acid (84), and Ca^{2+} ions (78), have clearly been shown to regulate the expression and properties of $\alpha 7$ nAChRs, suggesting that the developmental regulation of $\alpha 7$ nAChR expression may involve a complex set of mechanisms. These observations, along with the transient increased expression of $\alpha 7$ nAChRs in the developing CNS, are consistent with a putative role for this receptor in developmental processes.

A second series of studies have shown that $\alpha 7$ nAChRs may modulate the plasticity of neuronal circuitry. Early experiments in the toad optic tectum have demonstrated that α -BTX binding sites modulate the development and maintenance of retinotectal connections (79). Subsequent studies have shown that activation of $\alpha 7$ -type nAChRs leads to neurite retraction in both PC12 cells and isolated ciliary ganglion neurons, which is dependent on Ca^{2+} influx (80,81). Finally, in a more recent study, activation of $\alpha 7$ nAChRs on cultured neurons of rat olfactory bulb was reported to produce neuritic elongation (82). Although the results of these studies may appear contradictory, it is reasonable to predict that different neuronal cell types may possess diverse Ca^{2+} -sensitive biochemical signaling pathways and, thus, can react differently to increased intracellular Ca^{2+} (83). In addition, these processes may depend on the spatial distribution of $\alpha 7$ nAChRs on the cell membrane. Nevertheless, these observations indicate that $\alpha 7$ nAChRs are involved in establishing and regulating neuronal circuitry, probably through Ca^{2+} -dependent mechanisms.

Varying levels of intracellular Ca^{2+} can have graded effects on developing neurons, from altering gene expression to apoptotic cell death (4,83,98). Given the high Ca^{2+} permeability of $\alpha 7$ nAChRs (22,37,38,40), these channels are likely to influence many aspects of neuronal differentiation. Indeed, this has been implicated in a series of past and recent studies.

First, α -BTX-sensitive nAChRs have been reported to regulate mRNA production of neurotrophic growth factors in the adult rat hippocampus (85). Consistent with this observation, activation of α -BTX-sensitive nAChRs resulted in cell proliferation in a neuroendocrine cell line (86,87). Other studies have demonstrated a neuroprotective role for the $\alpha 7$ nAChR. Preactivation of $\alpha 7$ -type nAChRs protected rodent cultured neocortical neurons against *N*-methyl-D-aspartate (NMDA)-mediated glutamatergic excitotoxicity (88,89). Similar neuroprotective properties of $\alpha 7$ nAChRs were observed in models of ischemia and lesion-induced atrophy (89,90).

Finally, a series of studies have implicated $\alpha 7$ nAChRs in opposing processes of apoptosis and neurodegeneration. Blockade of α -BTX-sensitive nAChRs was previously reported to rescue embryonic chick motoneurons from naturally occurring cell death (91,92). More recently, activation of these receptors by nicotine was demonstrated to induce apoptotic cell death of rat hippocampal progenitor cells (93). Furthermore, an $\alpha 7$ nAChR mutation (L247T) resulting in a gain of channel function (94,95) has been demonstrated to cause neurodegenerative apoptosis in the *Caenorhabditis elegans* (96) and the developing murine nervous systems (97). These contrasting neuroprotective and neurodegenerative effects of $\alpha 7$ nAChRs on developing neurons can again be explained by the appearance of diverse biochemical signaling pathways, but also by the expression of Ca^{2+} -buffering mechanisms (93), which may change the effect of $\alpha 7$ activation from damaging to beneficial. Taken together, these data provide strong evidence that $\alpha 7$ nAChRs can mediate diverse effects on developing neurons ranging from local regulation of neurite growth cones to a more general modulation of cellular growth and differentiation.

Given the increasing evidence for the involvement of $\alpha 7$ nAChR in neuronal development and plasticity, it is surprising to learn that knockout of the $\alpha 7$ -subunit yields animals that survive normally with no apparent physical or neuroanatomical deficits (33). Moreover, these

mice show no obvious difference in the barrel formations within their somatosensory cortex (SS1), a region delineated by $\alpha 7$ nAChR expression during cortical development (70). However, preliminary studies have shown that $\alpha 7$ nAChR knockout mice display an anomalous synchronization on electroencephalography recordings (99), suggesting that loss of the $\alpha 7$ nAChR may cause more subtle phenotypic abnormalities in the intrinsic circuitry of the brain. In the absence of $\alpha 7$ nAChRs, other mechanisms may take over to help modulate intracellular Ca^{2+} levels (see $\alpha 7$ nAChRs and Glutamatergic Systems), resulting in apparently normal anatomical structures. Therefore, future studies may need to analyze these $\alpha 7$ -deficient mice on an ultrastructural level. Furthermore, the absence of the $\alpha 7$ nicotinic receptor subunit may only become evident when there is a compromise of neuronal function, such as in aging or some neurological disorder. This has recently been demonstrated for mice lacking the $\beta 2$ nicotinic receptor subunit (100). Additionally, other animal models expressing alterations of the $\alpha 7$ nAChR resulting in a "gain of function" (97) may help to elucidate the functional role of these receptors in the brain.

Early Cholinergic Circuitry

If $\alpha 7$ nAChRs are truly involved in modulating neuronal development, an established source of endogenous agonist, such as ACh, must be present within developing regions of the CNS that express $\alpha 7$ nAChRs. For this topic, we shall focus our discussion on the rodent sensory cortex, which expresses high levels of $\alpha 7$ nAChRs during brain development (70,71,73). The rodent cortex receives its primary cholinergic projection from ACh-containing neurons located in the basal forebrain (101,102). A transient expression of cholinergic neurons within rat sensory cortex has been reported to peak during the perinatal period (103). Interestingly, $\alpha 7$ mRNA and protein expression can also be observed during this time-point within the embryonic neocortex (70). Cortical ingrowth of basal forebrain afferents in the rodent has been

observed as early as postnatal d 0 (P0) (104,105). These tracing studies have indicated that the early postnatal development of basal forebrain fibers coincides temporally with a transient expression of $\alpha 7$ nAChRs in the sensory cortices (70,74). Whereas previous studies were not able to detect choline acetyltransferase-(ChAT) positive expression in these basal forebrain afferents before P5 (106), in a more recent study, ChAT-immunopositive axons were detected in the rat cerebral cortex at birth (107). This directly coincides with the first detectable expression of $\alpha 7$ nAChR mRNA and protein in the sensory cortex (74).

Although these results are in agreement, there is still an apparent discrepancy between the high levels of $\alpha 7$ nAChR expression and the relatively low levels of ChAT expression in the developing cortex. This becomes even more pronounced considering the high levels of acetylcholinesterase (AChE) activity in these developing cortical regions (108). However, recent investigations suggest that ACh may not be the primary activator of $\alpha 7$ nAChRs during brain development. These studies have demonstrated that choline, a precursor of ACh and a product of ACh hydrolysis by AChE, acts as an efficient and selective agonist of $\alpha 7$ nAChRs expressed in *Xenopus* oocytes and neuronal cell and slice cultures (62,109–111). Thus, high levels of choline, possibly owing to increased AChE expression, may play a prominent role in regulating $\alpha 7$ nAChR activity during development. Although the precise concentration and spatiotemporal distribution of choline in the brain remain to be determined, these findings indicate that $\alpha 7$ nAChRs may be functionally active during the critical period of neuronal growth and differentiation.

$\alpha 7$ nAChRs and Glutamatergic Systems

One issue that warrants further consideration is the consistent association of $\alpha 7$ nAChRs with glutamatergic transmission. This issue becomes particularly interesting given the elec-

trophysiological response profiles of $\alpha 7$ nAChRs and NMDA receptors (112). Although both receptor channels are highly permeable to Ca^{2+} , they are differentially regulated by Mg^{2+} and show opposing current-voltage relationships. ACh-induced $\alpha 7$ nAChR currents exhibit an inward rectification, which is dependent on the presence of intracellular Mg^{2+} (113,114). In contrast, the outward rectification of NMDA-evoked currents is dependent on the presence of extracellular Mg^{2+} (115). Thus, $\alpha 7$ nAChRs can mediate a substantial Ca^{2+} entry into neurons at resting or hyperpolarizing membrane potentials, whereas NMDA receptors gate Ca^{2+} entry into neurons at depolarized conditions. Additionally, NMDA receptor activation has been shown to induce an elevation in intracellular Mg^{2+} levels (116) that would ensure blockade of $\alpha 7$ nAChRs and prevent Ca^{2+} overload of neurons owing to simultaneous activation of both receptor types. These observations indicate complementary, nonoverlapping roles for $\alpha 7$ nAChRs and NMDA receptors in the regulation of intracellular Ca^{2+} concentrations, a concept that may be referred to as a “ying-and-yang” hypothesis. This hypothesis, initially put forth by Albuquerque and colleagues (112), suggests a mutual physiological interaction between the two transmitter systems that may serve to fine-tune cellular activity.

Perhaps one of the best examples of this putative interaction between cholinergic and glutamatergic systems can be seen in the developing rodent sensory cortex. Sensory regions of the cortex provide a useful model for analysis of synaptic plasticity, because they exhibit dynamic changes during early postnatal development. These cortical areas are innervated by afferents from the thalamus and basal forebrain, which provide the primary glutamatergic and cholinergic transmission to the cortex, respectively (102,117,118). Furthermore, these two transmitter pathways innervate the sensory cortex during a critical period of development (106,119–121). Studies have demonstrated that disruption of either the cholinergic (5,6,68) or glutamatergic (122,123)

pathways during this early period can have marked consequences on cortical development and plasticity.

Both $\alpha 7$ nAChRs and glutamatergic receptors are active and exhibit discrete patterns of expression in the sensory cortices at around the same postnatal period (45,70,71,73,124,125). In the developing rat SS1, studies have demonstrated that $\alpha 7$ nAChR expression on cortical neurons is tightly regulated by the ingrowing thalamocortical afferents (74) and that this regulation may be mediated by glutamate acting via NMDA receptors (126). Moreover, activation of presynaptic $\alpha 7$ nAChRs in the developing rat auditory cortex has recently been shown to enhance NMDA receptor-mediated glutamatergic transmission (45). These data imply that glutamatergic signals from ingrowing thalamocortical afferents cause an increase in postsynaptic and perhaps presynaptic $\alpha 7$ nAChR expression in the developing cortex. Together with NMDA receptors, postsynaptic $\alpha 7$ nAChRs may serve to regulate intracellular Ca^{2+} levels in cortical neurons (112) whereas presynaptic $\alpha 7$ nAChRs could serve as a feedback mechanism for modulating glutamatergic transmission (18). Such a scheme is presented in Fig. 1. In support of this overall hypothesis, several recent studies have demonstrated that activation of $\alpha 7$ nAChRs protected rat neuronal cultures from NMDA receptor-mediated excitotoxicity (88,89,127,128). These results indicate a close interaction between cholinergic and glutamatergic pathways, mediated by $\alpha 7$ nAChRs and NMDA receptors, that can have significant effects on neuronal plasticity throughout the developing and adult brain.

Clinical Implications

Over the past two decades, accumulating knowledge of nAChRs in vertebrates has led to the demonstration that alterations of these receptors may be responsible for a variety of familial disorders of the CNS and PNS (reviewed in ref. 129). Consequently, these

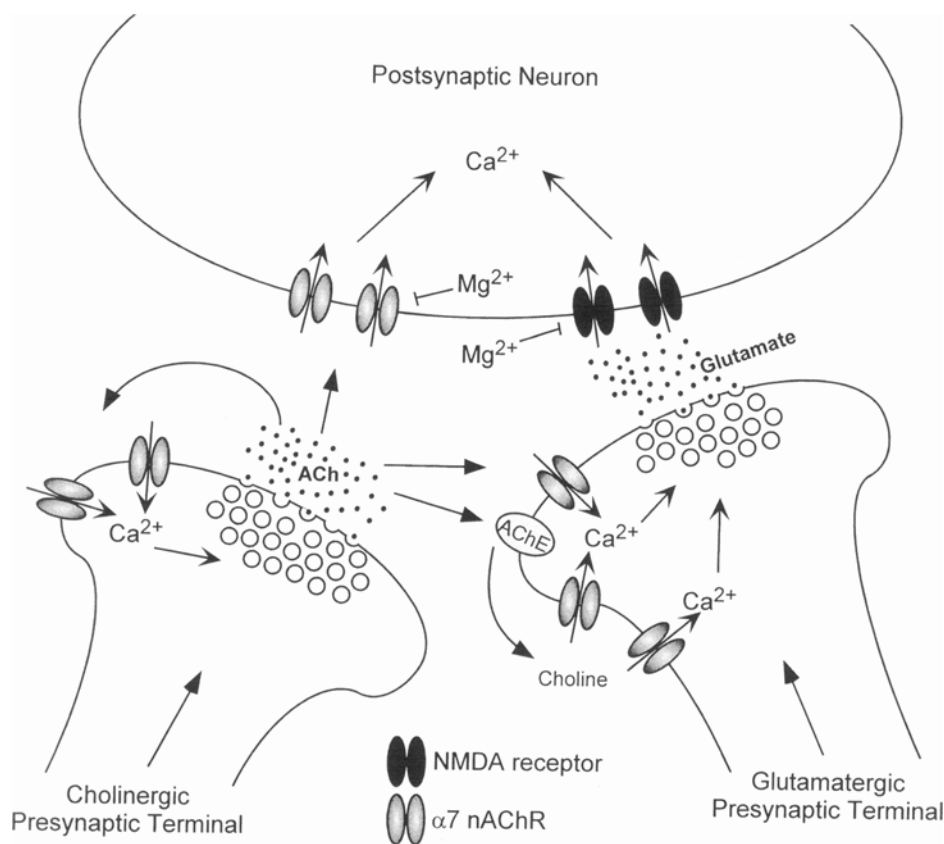


Fig. 1. Schematic diagram depicting the hypothetical roles for $\alpha 7$ nAChRs in the CNS. Presynaptic $\alpha 7$ nAChRs can modulate the release of neurotransmitters, whereas postsynaptic receptors can help regulate the levels of intracellular Ca^{2+} . Also presented is the putative interaction between the cholinergic and glutamatergic systems. Together, $\alpha 7$ nAChRs and NMDA receptors can regulate intracellular Ca^{2+} levels, with Mg^{2+} -mediated rectification as the limiting factor. This diagram represents a potential model for cholinergic/glutamatergic interactions throughout the brain. Adapted from Albuquerque et al. (17). Acetylcholinesterase, AChE.

receptors are also being considered as relevant targets for nicotinic therapies of brain disorders. Because of the potential for $\alpha 7$ nAChRs to modulate neuronal plasticity, their aberrant function may underlie many of these disorders. Indeed, the recent cloning of the human $\alpha 7$ nAChR (130) has generated increased interest in identifying disorders involving this receptor.

One topic that has attracted considerable interest in recent years is the problem of tobacco abuse. Although the underlying mechanisms that cause tobacco abuse are not well understood, accumulated evidence indicates that nicotine is the primary component of

tobacco that motivates continued use (29,30). Numerous studies have indicated that nicotine may have adverse effects on the developing CNS. The incidence of cigaret smoking among pregnant women is approx 20–50%, resulting in infant health problems, such as prematurity, low birthweight, and sudden infant death syndrome (131). In animal studies, prenatal nicotine exposure has been associated with abnormal neuronal maturation, defects in multiple neurotransmitter systems, and alterations in cognitive performance of the offspring (132–135). Moreover, fetal nicotine exposure has been observed to cause abnormal cortical

development (136). Anatomical analysis of the SS1 in exposed rat pups shows a reduction in cortical thickness and cell size, as well as decreased dendritic branching. Given the evidence presented, these observations implicate the involvement of nAChRs, including the $\alpha 7$ nAChR subtype, in mediating the adverse effects of nicotine in the developing brain, and warrant further investigation of this role.

Characterizing the functional role(s) of $\alpha 7$ nAChRs in the brain and the secondary mechanisms they could activate may also help in our understanding of a variety of neurological disorders. One such disorder where $\alpha 7$ nAChRs have received considerable attention is schizophrenia. This polygenetic disorder is partially characterized by an auditory gating deficit, an abnormal electrophysiological response to repeated auditory stimuli involving the hippocampus (137). Studies have shown that increased cigaret smoking in schizophrenic patients normalizes this aberrant trait (138). Recent investigations in both animal models and humans (139,140), in conjunction with familial linkage data (141), have indicated that decreased function of hippocampal $\alpha 7$ nAChRs could underlie the auditory gating deficit in schizophrenia. Although $\alpha 7$ -deficient mice appear to have normal sensorimotor gating (142), it is possible that what is important is not the number of nAChRs, but how they function. Thus, future studies may examine the gating responses of mice with mutations in the $\alpha 7$ gene that produce a receptor with altered function (97). Despite the usual appearance of characteristic symptoms in late adolescence or early adulthood, schizophrenia is increasingly seen as a deficit in brain growth and development, possibly involving a disruption of neuronal migration within the cortex (reviewed in 143–145). Therefore, the involvement of $\alpha 7$ nAChRs in neuronal growth and differentiation could allow them to influence this aspect of the pathogenesis of schizophrenia as well.

Finally, recent genetic studies in humans have found a significant link between the chromosomal region encompassing the $\alpha 7$ nAChR

gene and several different forms of idiopathic epilepsy (146,147). In addition, animal studies have identified polymorphisms in the $\alpha 7$ -subunit gene among strains of mice that are associated with sensitivity to nicotine-induced convulsions (148,149). Moreover, $\alpha 7$ -deficient mice show epileptiform-like wave patterns in the hippocampus (99). Together, these results suggest that $\alpha 7$ nAChRs may play a crucial role in regulating general neuronal activity.

Conclusions

A growing body of evidence has indicated that neuronal nAChRs may play a broader role than merely promoting fast cholinergic transmission. In particular, the $\alpha 7$ nAChR may have a distinct function in regulating synaptic plasticity. By modulating intracellular Ca^{2+} levels in discrete locations on neuronal cell bodies, dendrites, and terminal fields, these ligand-gated ion channels can regulate a myriad of physiological processes in the developing and adult brain. With the ongoing development of new pharmacological and molecular tools, the functional roles of this nicotinic receptor subtype may finally be elucidated.

Acknowledgments

The authors would like to thank Dr. M. DeBiasi for valuable comments on this manuscript. This work was supported by NIDA grants DA05947-01 and DA 10612.

References

1. Lipton S. A. and Kater S. B. (1989) Neurotransmitter regulation of neuronal outgrowth, plasticity and survival. *TINS* **12**, 265–270.
2. Leslie F. M. (1993) Neurotransmitters as neurotrophic factors, in *Neurotrophic Factors* (Loughlin S. E. and Fallon J. H., eds.) Academic, New York, pp. 565–598.
3. Levitt P., Harvey J. A., Friedman E., Simansky K., and Murphy E. H. (1997) New evidence for

- neurotransmitter influences on brain development. *Trends Neurosci.* **20**, 269–274.
4. Kater S. B. and Mills L. R. (1991) Regulation of growth cone behavior by calcium. *J. Neurosci.* **11**, 891–899.
 5. Bear M. F. and Singer W. (1986) Modulation of visual cortical plasticity by acetylcholine and noradrenaline. *Nature* **320**, 172–176.
 6. Hohmann C. F., Brooks A. R., and Coyle J. T. (1988) Neonatal lesions of the basal forebrain cholinergic neurons result in abnormal cortical development. *Dev. Brain Res.* **42**, 253–264.
 7. Juliano S. L., Ma W., and Elsin D. (1991) Cholinergic depletion prevents expansion of topographic maps in somatosensory cortex. *Proc. Natl. Acad. Sci. USA* **88**, 780–784.
 8. Juliano S. L. (1998) Mapping the sensory mosaic. *Science* **279**, 1653–1654.
 9. Gu Q. and Singer W. (1989) The role of muscarinic acetylcholine receptors in ocular dominance plasticity. *Experientia* **57**, 305–314.
 10. Role L. W. and Berg D. K. (1996) Nicotinic receptors in the development and modulation of CNS synapses. *Neuron* **16**, 1077–1085.
 11. Couturier S., Bertrand D., Matter J. M., Hernandez M. C., Bertrand S., Millar N., et al. (1990) A neuronal nicotinic acetylcholine receptor subunit ($\alpha 7$) is developmentally regulated and forms a homo-oligomeric channel blocked by α -BTX. *Neuron* **5**, 847–856.
 12. Clarke P. B. S. (1992) The fall and rise of neuronal α -bungarotoxin binding proteins. *TINS* **13**, 407–413.
 13. Deneris E. S., Connolly J., Rogers S. W., and Duvoisin R. (1991) Pharmacological and functional diversity of neuronal nicotinic acetylcholine receptors. *Trends Pharmacol. Sci.* **12**, 34–40.
 14. Sargent P. B. (1993) The diversity of neuronal nicotinic acetylcholine receptors. *Annu. Rev. Neurosci.* **16**, 403–443.
 15. Patrick J., Sequela P., Vernino S., Amador M., Luetje C., and Dani J. A. (1993) Functional diversity of neuronal nicotinic acetylcholine receptors. *Prog. Brain Res.* **98**, 113–120.
 16. McGehee D. S. and Role L. W. (1995) Physiological diversity of nicotinic acetylcholine receptors expressed by vertebrate neurons. *Annu. Rev. Physiol.* **57**, 521–546.
 17. Albuquerque E. X., Alkondon M., Pereira E. F. R., Castro N. G., Schrattenholz A., Barbosa C. T. F., et al. (1997) Properties of neuronal nicotinic acetylcholine receptors pharmacological characterization and modulation of synaptic function. *J. Pharm. Exp. Ther.* **280**, 1117–1136.
 18. Wonnacott S. (1997) Presynaptic nicotinic ACh receptors. *Trends Neurosci.* **20**, 92–98.
 19. Changeux J. P., Bertrand D., Corringer P. J., Dehaene S., Edelstein S., Lena C., et al. (1998) Brain nicotinic receptors: structure and regulation, role in learning and reinforcement. *Brain Res. Brain Res. Rev.* **26**, 198–216.
 20. Gerzanich V., Anand R., and Lindstrom J. (1994) Homomers of $\alpha 8$ and $\alpha 7$ subunits of nicotinic receptors exhibit similar channel but contrasting binding site properties. *Mol. Pharmacol.* **45**, 212–220.
 21. Elgoyhen A. B., Johnson D. S., Boulter J., Vetter D. E., and Heinemann S. (1994) $\alpha 9$: An acetylcholine receptor with novel pharmacological properties expressed in rat cochlear hair cells. *Cell* **79**, 705–715.
 22. Seguela P., Wadiche J., Dineley-Miller K., Dani J. A., and Patrick J. W. (1993) Molecular cloning, functional properties, and distribution of rat brain $\alpha 7$: a nicotinic cation channel highly permeable to calcium. *J. Neurosci.* **13**, 596–604.
 23. Wada E., Wada K., Boulter J., Deneris E., Heinemann S., Patrick J., et al. (1989) Distribution of alpha 2, alpha 3, alpha 4, and beta 2 neuronal nicotinic receptor subunit mRNAs in the central nervous system: a hybridization histochemical study in the rat. *J. Comp. Neurol.* **284**, 314–335.
 24. Flores C. M., Rogers S. W., Pabreza L. A., Wolfe B. B., and Kellar K. J. (1992) A subtype of nicotinic cholinergic receptor in rat brain is composed of $\alpha 4$ and $\beta 2$ subunits and is up-regulated by chronic nicotine treatment. *Mol. Pharmacol.* **41**, 31–37.
 25. Alkondon M. and Albuquerque E. X. (1993) Diversity of nicotinic acetylcholine receptors in rat hippocampal neurons. I. Pharmacological and functional evidence for distinct structural subtypes. *J. Pharmacol. Exp. Ther.* **265**, 1455–1473.
 26. Zoli M., Lena C., Picciotto M. R., and Changeux J. P. (1998) Identification of four classes of brain nicotinic receptors using $\beta 2$ mutant mice. *J. Neurosci.* **18**, 4461–4472.
 27. Charpentier E., Barneoud P., Moser P., Besnard F., and Sgard F. (1998) Nicotinic acetylcholine subunit mRNA expression in dopaminergic neurons of the rat substantia nigra and ventral tegmental area. *Neuroreport* **9**, 3097–3101.

28. Sorenson E. M., Shiroyama T., and Kitai S. T. (1998) Postsynaptic nicotinic receptors on dopaminergic neurons in the substantia nigra pars compacta of the rat. *Neuroscience* **87**, 659–673.
29. Stolerman I. P. and Shoaib M. (1991) The neurobiology of tobacco addiction. *TiPS* **12**, 467–473.
30. Dani J. A. and Heinemann S. (1996) Molecular and cellular aspects of nicotine abuse. *Neuron* **16**, 905–908.
31. Picciotto M. R., Zoli M., Rimondini R., Lena C., Marubio L. M., Pich E. M., et al. (1998) Acetylcholine receptors containing the $\beta 2$ subunit are involved in the reinforcing properties of nicotine. *Nature* **391**, 173–177.
32. Chen D. and Patrick J. W. (1997) The α -bungarotoxin-binding nicotinic acetylcholine receptor from rat brain contains only the $\alpha 7$ subunit. *J. Biol. Chem.* **272**, 24,024–24,029.
33. Orr-Urtreger A., Goldner F. M., Saeki M., Lorenzo I., Goldberg L., De Biasi M., et al. (1997) Mice deficient in the $\alpha 7$ neuronal nicotinic acetylcholine receptor lack α -bungarotoxin binding sites and hippocampal fast nicotinic currents. *J. Neurosci.* **17**, 9165–9171.
34. Anand R., Peng X., and Lindstrom J. (1993) Homomeric and native $\alpha 7$ acetylcholine receptors exhibit remarkably similar but non-identical pharmacological properties, suggesting that the native receptor is a heteromeric protein complex. *FEBS* **327**, 241–246.
35. Gotti C., Hanke W., Maury K., Moretti M., Ballivet M., Clementi F., and Bertrand D. (1994) Pharmacology and biophysical properties of the $\alpha 7$ and $\alpha 7$ - $\alpha 8$ α -bungarotoxin receptor subtypes immunopurified from the chick optic lobe. *Eur. J. Neurosci.* **6**, 1281–1291.
36. Yu C. R. and Role L. W. (1998) Functional contribution of the $\alpha 5$ subunit to neuronal nicotinic channels expressed by chick sympathetic ganglion neurones. *J. Physiol. (Lond.)* **509**, 667–681.
37. Zhang Z.-w., Vijayaraghavan S., and Berg D. K. (1994) Neuronal acetylcholine receptors that bind α -bungarotoxin with high affinity function as ligand-gated ion channels. *Neuron* **12**, 167–177.
38. Vijayaraghavan S., Pugh P. C., Zhang Z. W., Rathouz M. M., and Berg D. K. (1992) Nicotinic receptors that bind α -bungarotoxin on neurons raise intracellular free Ca^{2+} . *Neuron* **8**, 353–362.
39. Barrantes G. E., Murphy C. T., Westwick J., and Wonnacott S. (1995) Nicotine increases intracellular calcium in rat hippocampal neurons via voltage-gated calcium channels. *Neurosci Lett.* **196**, 101–104.
40. Castro N. G. and Albuquerque E. X. (1995) α -Bungarotoxin-sensitive hippocampal nicotinic receptor channel has a high calcium permeability. *Biophys. J.* **68**, 516–524.
41. Gray T., Rajan A. S., Radcliffe K. A., Yakehiro Y., and Dani J. A. (1996) Hippocampal synaptic transmission enhanced by low concentrations of nicotine. *Nature* **383**, 713–716.
42. McGehee D. S., Heath M. J., Gelber S., Devay P., and Role L. W. (1995) Nicotine enhancement of fast excitatory synaptic transmission in CNS by presynaptic receptors. *Science* **269**, 1692–1696.
43. Guo J. Z., Tredway T. L., and Chiappinelli V. A. (1998) Glutamate and GABA release are enhanced by different subtypes of presynaptic nicotinic receptors in the lateral geniculate nucleus. *J. Neurosci.* **18**, 1963–1969.
44. Alkondon M., Rocha E. S., Maelicke A., and Albuquerque E. X. (1996) Diversity of nicotinic acetylcholine receptors in rat brain. V. α -Bungarotoxin-sensitive nicotinic receptors in olfactory bulb neurons and presynaptic modulation of glutamate release. *J. Pharmacol. Exp. Ther.* **278**, 1460–1471.
45. Aramakis V. B. and Metherate R. (1998) Nicotine selectively enhances NMDA receptor-mediated synaptic transmission during postnatal development in sensory neocortex. *J. Neurosci.* **18**, 8485–8495.
46. Pidoplichko V. I., DeBiasi M., Williams J. T., and Dani J. A. (1997) Nicotine activates and desensitizes midbrain dopamine neurons. *Nature* **390**, 401–404.
47. Zarei M. M., Radcliffe K. A., Chen D., Patrick J. W., and Dani J. A. (1999) Distribution of nicotinic acetylcholine receptor $\alpha 7$ and $\beta 2$ subunits on cultured hippocampal neurons. *Neurosci.* **88**, 755–764.
48. Schilstrom B., Svensson H. M., Svensson T. H., and Nomikos G. G. (1998) Nicotine and food induced dopamine release in the nucleus accumbens of the rat: putative role of $\alpha 7$ nicotinic receptors in the ventral tegmental area. *Neuroscience* **85**, 1005–1009.
49. Li X., Rainnie D. G., McCarley R. W., and Greene R. W. (1998) Presynaptic nicotinic receptors facilitate monoaminergic transmission. *J. Neurosci.* **18**, 1904–1912.

50. Liang S.-D. and Vizi E. S. (1997) Positive feedback modulation of acetylcholine release from isolated rat superior cervical ganglion. *J. Pharm. Exp. Ther.* **280**, 650–655.
51. Radcliffe K. A., Fisher J. L., Gray R., and Dani J. A. (1999) Nicotinic modulation of glutamate and GABA synaptic transmission in hippocampal neurons. *Ann. NY Acad. Sci.* **868**, 591–610.
52. Alkondon M., Pereira E. F., Barbosa C. T., and Albuquerque E. X. (1997) Neuronal nicotinic acetylcholine receptor activation modulates gamma-aminobutyric acid release from CA1 neurons of rat hippocampal slices. *J. Pharmacol. Exp. Ther.* **283**, 1396–1411.
53. Alkondon M., Pereira E. F., and Albuquerque E. X. (1998) α -bungarotoxin- and methyllycaconitine-sensitive nicotinic receptors mediate fast synaptic transmission in interneurons of rat hippocampal slices. *Brain Res.* **810**, 257–263.
54. Frazier C. J., Buhler A. V., Weiner J. L., and Dunwiddie T. V. (1998) Synaptic potentials mediated via α -bungarotoxin-sensitive nicotinic acetylcholine receptors in rat hippocampal interneurons. *J. Neurosci.* **18**, 8228–8235.
55. Frazier C. J., Rollins Y. D., Breese C. R., Leonard S., Freedman R., and Dunwiddie T. V. (1998) Acetylcholine activates an α -bungarotoxin-sensitive nicotinic current in rat hippocampal interneurons, but not pyramidal cells. *J. Neurosci.* **18**, 1187–1195.
56. Corriveau R. A. and Berg D. K. (1993) Coexpression of multiple acetylcholine receptor genes in neurons: Quantification of transcripts during development. *J. Neurosci.* **13**, 2662–2671.
57. Jacob M. H. and Berg D. K. (1983) The ultrastructural localization of α -bungarotoxin binding sites in relation to synapses on chick ciliary ganglion neurons. *J. Neurosci.* **3**, 260–271.
58. Horch H. L. and Sargent P. B. (1995) Perisynaptic surface distribution of multiple classes of nicotinic acetylcholine receptors on neurons in the chicken ciliary ganglion. *J. Neurosci.* **15**, 7778–7795.
59. Zhang Z. W., Coggan J. S., and Berg D. K. (1996) Synaptic currents generated by neuronal acetylcholine receptors sensitive to α -bungarotoxin. *Neuron* **17**, 1231–1240.
60. Ullian E. M., McIntosh J. M., and Sargent P. B. (1997) Rapid synaptic transmission in the avian ciliary ganglion is mediated by two distinct classes of nicotinic receptors. *J. Neurosci.* **17**, 7210–7219.
61. Chang K. T. and Berg D. K. (1999) Nicotinic acetylcholine receptors containing $\alpha 7$ subunits are required for reliable synaptic transmission *in situ*. *J. Neurosci.* **19**, 3701–3710.
62. Alkondon M., Pereira E. F., Eisenberg H. M., and Albuquerque E. X. (1999) Choline and selective antagonists identify two subtypes of nicotinic acetylcholine receptors that modulate GABA release from CA1 interneurons in rat hippocampal slices. *J. Neurosci.* **19**, 2693–2705.
63. Hunter B. E., de Fiebre C. M., Papke R. L., Kem W. R., and Meyer E. M. (1994) A novel nicotinic agonist facilitates induction of long-term potentiation in the rat hippocampus. *Neurosci. Lett.* **168**, 130–134.
64. Radcliffe K. A. and Dani J. A. (1998) Nicotinic stimulation produces multiple forms of increased glutamatergic synaptic transmission. *J. Neurosci.* **18**, 7075–7083.
65. Baskerville K. A., Schweitzer J. B., and Herron, P. (1997) Effects of cholinergic depletion on experience-dependent plasticity in the cortex of the rat. *Neuroscience* **80**, 1159–1169.
66. Kilgard M. P. and Merzenich M. M. (1998) Cortical map reorganization enabled by nucleus basalis activity. *Science* **279**, 1714–1718.
67. Sachdev R. N., Lu S. M., Wiley R. G., and Ebner F. F. (1998) Role of the basal forebrain cholinergic projection in somatosensory cortical plasticity. *J. Neurophysiol.* **79**, 3216–3228.
68. Zhu X. O. and Waite P. M. (1998) Cholinergic depletion reduces plasticity of barrel field cortex. *Cereb. Cortex* **8**, 63–72.
69. Wang G. K. and Schmidt J. (1976) Receptors for α -bungarotoxin in the developing visual system of the chick. *Brain Res.* **114**, 524–529.
70. Broide R. S., O'Connor L. T., Smith M. A., Smith J. A., and Leslie F. M. (1995) Developmental expression of $\alpha 7$ neuronal nicotinic receptor messenger RNA in rat sensory cortex and thalamus. *Neuroscience* **67**, 83–94.
71. Fuchs J. L. (1989) [125I] α -bungarotoxin binding marks primary sensory area developing rat neocortex. *Brain Res.* **501**, 223–234.
72. Fiedler E. P., Marks M. J., and Collins A. C. (1990) Postnatal development of two nicotinic cholinergic receptors in seven mouse brain regions. *Int. J. Dev. Neurosci.* **8**, 533–540.
73. Bina K. G., Guzman P., Broide R. S., Leslie F. M., Smith M. A., and O'Dowd, D. K. (1995) Localization of $\alpha 7$ nicotinic receptor subunit mRNA and α -bungarotoxin binding sites in

- developing mouse somatosensory thalamo-cortical system. *J. Comp. Neurol.* **363**, 321–332.
74. Broide R. S., Robertson R. T., and Leslie F. M. (1996) Regulation of $\alpha 7$ nicotinic acetylcholine receptors in the developing rat somatosensory cortex by thalamocortical afferents. *J. Neurosci.* **16**, 2956–2971.
 75. Halvorsen S. W. and Berg D. K. (1989) Specific down-regulation of the α -bungarotoxin binding component on chick autonomic neurons by ciliary neuronotrophic factor. *J. Neurosci.* **9**, 3673–3680.
 76. Geertsen S., Trifaro J. M., and Quik M. (1992) Phorbol esters and K^+ up-regulate α -bungarotoxin binding sites in cultured chromaffin cells through a related mechanism. *Neurosci. Lett.* **148**, 207–210.
 77. De Koninck P. and Cooper E. (1995) Differential regulation of neuronal nicotinic ACh receptor subunit genes in cultured neonatal rat sympathetic neurons: specific induction of $\alpha 7$ by membrane depolarization through a Ca^{2+} /calmodulin-dependent kinase pathway. *J. Neurosci.* **15**, 7966–7978.
 78. Ospina J. A., Broide R. S., Acevedo D., Robertson R. T., and Leslie F. M. (1998) Calcium regulation of agonist binding to $\alpha 7$ -type nicotinic acetylcholine receptors in adult and fetal rat hippocampus. *J. Neurochem.* **70**, 1061–1068.
 79. Freeman J. A. (1977) Possible regulatory function of acetylcholine receptor in maintenance of retinotectal synapses. *Nature* **269**, 218–222.
 80. Chan J. and Quik M. (1993) A role for the nicotinic α -bungarotoxin receptor in neurite outgrowth in PC12 cells. *Neuroscience* **56**, 441–451.
 81. Pugh P. C. and Berg D. K. (1994) Neuronal acetylcholine receptors that bind α -bungarotoxin mediate neurite retraction in a calcium-dependent manner. *J. Neurosci.* **14**, 889–896.
 82. Coronas V., Durand M., Jourdan F., and Quirion R. (1998) Morphogenic effects of acetylcholine in rat olfactory bulb primary cultures. *Soc. Neurosci. Abstr.* **24**, 1338.
 83. Ghosh A. and Greenberg M. E. (1995) Calcium signaling in neurons: molecular mechanisms and cellular consequences. *Science* **268**, 239–247.
 84. Vijayaraghavan S., Huang B., Blumenthal E. M., and Berg D. K. (1995) Arachidonic acid as a possible negative feedback inhibitor of nicotinic acetylcholine receptors on neurons. *J. Neurosci.* **15**, 3679–3687.
 85. Freedman R., Wetmore C., Stromberg I., Leonard S., and Olson L. (1993) α -bungarotoxin binding to hippocampal interneurons: immunocytochemical characterization and effects on growth factor expression. *J. Neurosci.* **13**, 1965–1975.
 86. Codignola A., Tarroni P., Cattaneo M. G., Vicentini L. M., Clementi F. and Sher E. (1994) Serotonin release and cell proliferation are under the control of alpha-bungarotoxin-sensitive nicotinic receptors in small-cell lung carcinoma cell lines. *FEBS Lett.* **342**, 286–290.
 87. Quik M., Chan J., and Patrick J. (1994) α -Bungarotoxin blocks the nicotinic receptor mediated increase in cell number in a neuroendocrine cell line. *Brain Res.* **655**, 161–167.
 88. Carlson N. G., Bacchi A., Rogers S. W., and Gahring L. C. (1998) Nicotine blocks TNF α mediated neuroprotection to NMDA by an α -bungarotoxin-sensitive pathway. *J. Neurobiol.* **35**, 29–36.
 89. Shimohama S., Greenwald D. L., Shafron D. H., Akaika A., Maeda T., Kaneko S., et al. (1998) Nicotinic $\alpha 7$ receptors protect against glutamate neurotoxicity and neuronal ischemic damage. *Brain Res* **779**, 359–363.
 90. Meyer E. M., King M. A., and Meyers C. (1998) Neuroprotective effects of 2, 4-dimethoxybenzylidene anabaseine (DMXB) and tetrahydroaminoacridine (THA) in neocortices of nucleus basalis lesioned rats. *Brain Res.* **786**, 252–254.
 91. Renshaw G., Rigby P., Self G., Lamb A., and Goldie R. (1993) Exogenously administered α -bungarotoxin binds to embryonic chick spinal cord: implications for the toxin-induced arrest of naturally occurring motoneuron death. *Neuroscience* **53**, 1163–1172.
 92. Hory-Lee F. and Frank E. (1995) The nicotinic blocking agents *d*-tubocurarine and α -bungarotoxin save motoneurons from naturally occurring death in the absence of neuromuscular blockade. *J. Neurosci.* **15**, 6453–6460.
 93. Berger F., Gage F. H., and Vijayaraghavan S. (1998) Nicotinic receptor-induced apoptotic cell death of hippocampal progenitor cells. *J. Neurosci.* **18**, 6871–6881.
 94. Revah F., Bertrand D., Galzi J. L., Devillers-Thiery A., Mulle C., Hussy N., et al. (1991) Mutations in the channel domain alter desensitization of a neuronal nicotinic receptor. *Nature* **353**, 846–849.

95. Bertrand D., Devillers-Thiery A., Revah F., Galzi J. L., Hussy N., Mulle C., et al. (1992) Unconventional pharmacology of a neuronal nicotinic receptor mutated in the channel domain. *Proc. Natl. Acad. Sci. USA* **89**, 1261–1265.
96. Treinin M. and Chalfie M. (1995) A mutated acetylcholine receptor subunit causes neuronal degeneration in *C. elegans*. *Neuron* **14**, 871–877.
97. Broide R. S., Orr-Urtreger A., Dang H., Kasten M. R., Dani J. A., Beaudet A. L., et al. (1998) Mice homozygous for the $\alpha 7$ L247T nicotinic acetylcholine receptor mutation show increased neuronal apoptosis and die within 24 hours of birth. *Soc. Neurosci. Abstr.* **24**, 834.
98. Orrenius S. and Nicotera P. (1994) The calcium ion and cell death. *J. Neural. Transm.* **43**, 1–11.
99. Orr-Urtreger A., Noebels J. L., Goldner F. M., Patrick J., and Beaudet A. L. (1996) A novel hypersynchronous neocortical EEG phenotype in mice deficient in the neuronal nicotinic acetylcholine receptor (nAChRs) $\alpha 7$ subunit gene. *Am. J. Hum. Genet.* **59**, A53.
100. Zoli M., Picciotto M. R., Ferrari R., Cocchi D., and Changeux J. P. (1999) Increased neurodegeneration during ageing in mice lacking high-affinity nicotine receptors. *EMBO J.* **18**, 1235–1244.
101. Semba K. and Fibiger H. C. (1989) Organization of central cholinergic systems. *Prog. Brain Res.* **79**, 37–63.
102. Woolf N. J. (1991) Cholinergic systems in mammalian brain and spinal cord. *Prog. Neurobiol.* **37**, 475–524.
103. Dori I. and Parnavelas J. G. (1989) The cholinergic innervation of the rat cerebral cortex shows two distinct phases in development. *Exp. Brain Res.* **76**, 417–423.
104. Dinopoulos A., Eadie L. A., Dori I., and Parnavelas J. G. (1989) The development of basal forebrain projections to the rat visual cortex. *Exp. Brain Res.* **76**, 563–571.
105. Calarco C. A. and Robertson R. T. (1995) Development of basal forebrain projections to visual cortex: Dil studies in rat. *J. Comp. Neurol.* **354**, 608–626.
106. Kiss J. and Patel A. J. (1992) Development of the cholinergic fibres innervating the cerebral cortex of the rat. *Int. J. Dev. Neurosci.* **10**, 153–170.
107. Mechawar N. and Descarries L. (1998) Early postnatal development of the cholinergic innervation in the rat cerebral cortex. *Soc. Neurosci. Abstr.* **24**, 1338.
108. Robertson R. T. (1987) A morphogenic role for transiently expressed acetylcholinesterase in developing thalamocortical systems. *Neurosci. Lett.* **75**, 259–264.
109. Papke R. L., Bencherif M., and Lippiello P. (1996) An evaluation of neuronal nicotinic acetylcholine receptor activation by quaternary nitrogen compounds indicates that choline is selective for the $\alpha 7$ subtype. *Neurosci. Lett.* **213**, 201–204.
110. Alkondon M., Pereira E. F., Cortes W. S., Maelicke A., and Albuquerque E. X. (1997) Choline is a selective agonist of $\alpha 7$ nicotinic acetylcholine receptors in the rat brain neurons. *Eur. J. Neurosci.* **9**, 2734–2742.
111. Meyer E. M., Tay E. T., Zoltewicz J. A., Meyers C., King M. A., Papke R. L., et al. (1998) Neuroprotective and memory-related actions of novel alpha-7 nicotinic agents with different mixed agonist/antagonist properties. *J. Pharmacol. Exp. Ther.* **284**, 1026–1032.
112. Albuquerque E. X., Pereira E. F. R., Castro N. G., and Alkondon M. (1995) Neuronal nicotinic receptors: function, modulation and structure. *The Neurosciences* **7**, 91–101.
113. Alkondon M., Reinhardt S., Lobron C., Hermesen B., Maelicke A., and Albuquerque E. X. (1994) Diversity of nicotinic acetylcholine receptors in rat hippocampal neurons. II. The rundown and inward rectification of agonist-elicited whole-cell currents and identification of receptor subunits by in situ hybridization. *J. Pharmacol. Exp. Ther.* **271**, 494–506.
114. Forster I. and Bertrand D. (1995) Inward rectification of neuronal nicotinic acetylcholine receptors investigated by using the homomeric $\alpha 7$ receptor. *Proc. R. Soc. Lond. B. Biol. Sci.* **260**, 139–148.
115. Mayer M. L., Westbrook G. L., and Guthrie P. B. (1984) Voltage-dependent block by Mg^{2+} of NMDA responses in spinal cord neurones. *Nature* **309**, 261–263.
116. Brocard J. B., Rajdev S., and Reynolds I. J. (1993) Glutamate-induced increases in intracellular free Mg^{2+} in cultured cortical neurons. *Neuron* **11**, 751–757.
117. Kharazia V. N. and Weinberg R. J. (1993) Glutamate in terminals of thalamocortical fibers in rat somatic sensory cortex. *Neurosci. Letts.* **157**, 162–166.

118. Gil Z. and Amitai Y. (1996) Adult thalamocortical transmission involves both NMDA and non-NMDA receptors. *J. Neurophys.* **76**, 2547–2554.
119. Hohmann C. F. and Ebner F. F. (1985) Development of cholinergic markers in mouse forebrain. I. Choline acetyltransferase enzyme activity and acetylcholinesterase histochemistry. *Dev. Brain Res.* **23**, 225–241.
120. Catalano S. M., Robertson R. T., and Killackey H. P. (1991) Early ingrowth of thalamocortical afferents to the neocortex of the prenatal rat. *Proc. Natl. Acad. Sci. USA* **88**, 2999–3003.
121. Miller B., Chou L., and Finlay B. L. (1993) The early development of thalamocortical and corticothalamic projections. *J. Comp. Neurol.* **335**, 16–41.
122. Schlaggar B. L., Fox K., and O'Leary D. D. M. (1993) Postsynaptic control of plasticity in developing somatosensory cortex. *Nature* **364**, 623–626.
123. Fox K., Schlaggar B. L., Glazewski S., and O'Leary D. D. M. (1996) Glutamate receptor blockade at cortical synapses disrupts development of thalamocortical and columnar organization in somatosensory cortex. *Proc. Natl. Acad. Sci. USA* **93**, 5584–5589.
124. Agmon A. and O'Dowd D. K. (1992) NMDA receptor-mediated currents are prominent in the thalamocortical synaptic response before maturation of inhibition. *J. Neurophys.* **68**, 345–349.
125. Blue M. E. and Johnston M. V. (1995) The ontogeny of glutamate receptors in rat barrel field cortex. *Dev. Brain Res.* **84**, 11–25.
126. Jensen J. J., Winzer-Serhan U. H., and Leslie F. M. (1997) Glial regulation of $\alpha 7$ -type nicotinic acetylcholine receptor expression in cultured rat cortical neurons. *J. Neurochem.* **68**, 112–120.
127. Kaneko S., Maeda T., Kume T., Kochiyama H., Akaike A., Shimohama S., et al. (1997) Nicotine protects cultured cortical neurons against glutamate-induced cytotoxicity via $\alpha 7$ -neuronal receptors and neuronal CNS receptors. *Brain Res.* **765**, 135–140.
128. Minana M. D., Montoliu C., Llansola M., Grisolia S., and Felipo V. (1998) Nicotine prevents glutamate-induced proteolysis of the microtubule-associated protein MAP-2 and glutamate neurotoxicity in primary cultures of cerebellar neurons. *Neuropharmacology* **37**, 847–857.
129. Lena C. and Changeux J. P. (1997) Pathological mutations of nicotinic receptors and nicotine-based therapies for brain disorders. *Curr. Opinion Neurobiol.* **7**, 674–682.
130. Peng X., Katz M., Gerzanich V., Anand R., and Lindstrom J. (1994) Human $\alpha 7$ acetylcholine receptor: cloning of the $\alpha 7$ subunit from the SH-SY5Y cell line and determination of pharmacological properties of native receptors and functional $\alpha 7$ homomers expressed in *Xenopus* oocytes. *Mol. Pharmacol.* **45**, 546–554.
131. Scheibmeir M. and O'Connell K. A. (1997) In harm's way: childbearing women and nicotine. *J. Obstet. Gynecol. Neonatal Nurs.* **26**, 477–484.
132. Navarro H. A., Seidler F. J., Eylers J. P., Baker F. E., Dobbins S. S., Lappi S. E., et al. (1989) Effects of prenatal nicotine exposure on development of central and peripheral cholinergic neurotransmitter systems. Evidence for cholinergic trophic influences in developing brain. *J. Pharmacol. Exp. Ther.* **251**, 894–900.
133. Navarro H. A., Seidler F. J., Schwartz R. D., Baker F. E., Dobbins S. S., and Slotkin T. A. (1989) Prenatal exposure to nicotine impairs nervous system development at a dose which does not affect viability or growth. *Brain Res. Bull.* **23**, 187–192.
134. Zahalka E. A., Seidler F. J., Lappi S. E., McCook E. C., Yanai J., and Slotkin T. A. (1992) Deficits in development of central cholinergic pathways caused by fetal nicotine exposure: differential effects on choline acetyltransferase activity and $[3H]$ hemicholinium-3 binding. *Neurotoxicol. Teratol.* **14**, 375–382.
135. Levin E. D., Briggs S. J., Channelle Christopher N., and Rose, J. E. (1993) Prenatal nicotine exposure and cognitive performance in rats. *Neurotoxicol. Teratol.* **15**, 251–260.
136. Roy T. S. and Sabherwal U. (1994) Effects of prenatal nicotine exposure on the morphogenesis of somatosensory cortex. *Neurotoxicol. Teratol.* **16**, 411–421.
137. Braff D. L. and Geyer M. A. (1990) Sensorimotor gating and schizophrenia. *Arch. Gen. Psychiatry* **47**, 181–188.
138. Adler L. E., Hoffer L. D., Wiser A., and Freedman R. (1993) Normalization of auditory physiology by cigarette smoking in schizophrenic patients. *Am. J. Psychiatry* **150**, 1856–1861.
139. Stevens K. E., Freedman R., Collins A. C., Hall M., Leonard S., Marks M. J., et al. (1996)

- Genetic correlation of inhibitory gating of hippocampal auditory evoked response and α -bungarotoxin-binding nicotinic cholinergic receptors in inbred mouse strains. *Neuropsychopharmacology* **15**, 152–162.
140. Adler L. E., Olincy A., Waldo M., Harris J. G., Griffith J., Stevens K., et al. (1998) Schizophrenia, sensory gating, and nicotinic receptors. *Schizophr Bull.* **24**, 189–202.
 141. Freedman R., Coon H., Myles-Worsley M., Orr-Urtreger A., Olincy A., Davis A., et al. (1997) Linkage of a neurophysiological deficit in schizophrenia to a chromosome 15 locus. *Proc. Natl. Acad. Sci. USA* **94**, 587–592.
 142. Paylor R., Nguyen M., Crawley J. N., Patrick J., Beaudet A., and Orr-Urtreger A. (1998) $\alpha 7$ nicotinic receptor subunits are not necessary for hippocampal-dependent learning or sensorimotor gating: a behavioral characterization of *acra7*-deficient mice. *Learning & Memory* **5**, 302–316.
 143. Jones E. G. (1997) Cortical development and thalamic pathology in schizophrenia. *Schizophr. Bull.* **23**, 483–501.
 144. Woolf C. M. (1997) Does the genotype for schizophrenia often remain unexpressed because of canalization and stochastic events during development. *Psychological Med.* **27**, 659–668.
 145. Raedler T. J., Knable M. B., and Weinberger D. R. (1998) Schizophrenia as a developmental disorder of the cerebral cortex. *Curr. Opinion Neurobiol.* **8**, 157–161.
 146. Elmslie F. V., Rees M., Williamson M. P., Kerr M., Kjeldsen M. J., Pang K. A., et al. (1997) Genetic mapping of a major susceptibility locus for juvenile myoclonic epilepsy on chromosome 15q. *Hum. Mol. Genet.* **6**, 1329–1334.
 147. Neubauer B. A., Fielder B., Himmelein B., Kampfer F., Labker U., Schwabe G., et al. (1998) Centrottemporal spikes in families with rolandic epilepsy: Linkage to chromosome 15q14. *Neurology* **51**, 1608–1612.
 148. Stitzel J. A., Farnham D. A., and Collins A. C. (1996) Linkage of strain-specific nicotinic receptor alpha 7 subunit restriction fragment length polymorphisms with levels of alpha-bungarotoxin binding in brain. *Brain Res. Mol. Brain. Res.* **43**, 30–40.
 149. Stitzel J. A., Blanchette J. M., and Collins A. C. (1998) Sensitivity to the seizure-inducing effects of nicotine is associated with strain-specific variants of the alpha 5 and alpha 7 nicotinic receptor subunit genes. *J. Pharmacol. Exp. Ther.* **284**, 1104–1111.